Post-transcriptional Regulation of Tetraspanins CD151 and CD9 by micro-RNAs in Prostate and Breast Cancers

Danielle Bond

BBiomed Sci (Hons)

Thesis submitted in fulfilment of the requirements for obtaining the degree of

Doctor of Philosophy (Medical Biochemistry)

March 2015

Declaration

The thesis contains no material which has been accepted for the award of any other degree or diploma in any university or other tertiary institution and, to the best of my knowledge and belief, contains no material previously published or written by another person, except where due reference has been made in the text. I give consent to the final version of my thesis being made available worldwide when deposited in the University's Digital Repository**, subject to the provisions of the Copyright Act 1968. **Unless an Embargo has been approved for a determined period.

.....

Danielle Bond

Acknowledgements

Completing a PhD has been an amazing experience, with many ups and not so many downs. I would like to firstly thank my mum, dad and all of my family for all the support and encouragement they have given me over the past four years especially. Without their support I don't think this achievement would have been possible.

Next I would like to thank my primary supervisor Dr Jude Weidenhofer for being a fantastic supervisor, mentor and friend. This thesis is testament to all the hard work and dedication you have put into shaping me into a young early career researcher. I can't thank you enough for all your help and advice.

I would also like to thank my co-supervisors Dr Murray Cairns and Prof. Leonie Ashman for all their help and advice throughout my PhD.

I want to specially thank Crystal, a fellow PhD student and friend, for keeping me sane and for all the fun times we have shared along the way. To Helen, Josh, Elham, Kristen, Hayley, Nikita and Rich, for making coming to 'work' relaxing and fun and for always being around if I needed a hand with anything.

Last but not least, I would like to thank Dr Kathryn Skelding, Prof. Rick Thorne and Dr Charles De Bock, for always giving me great advice. Also I would like to thank members of the Cairns lab, particularly Belinda Goldie, Dr Adam Carroll and Dr Natalie Beveridge for teaching me miRNA techniques and always being available when I have had questions regarding miRNAs.

Table of Contents

RNAs in Prostate and Breast Cancers	
Declaration	2
Acknowledgements	3
Table of Contents	4
List of figures	
List of tables	
Abstract	
Publications and conference abstracts arising from this thesis	
Abbreviations	
Chapter 1: Literature Review	
1.1 Breast and prostate cancer statistics	
1.2 The pathogenesis of breast and prostate cancer	
1.4.1 Tetraspanin CD151	
1.4.2 CD151 and prostate cancer	
1.4.3 CD151 and breast cancer	
1.4.4 Tetraspanin CD9	
1.4.5 CD9 and prostate cancer	
1.4.6 CD9 and breast cancer	
1.5 Regulation of tetraspanin expression	
1.5.1 Regulation of CD9 expression	
1.5.2 Regulation of CD151 expression	35
1.5.3 Post-transcriptional silencing by micro-RNAs	37
1.6 Rationale, Aims & Hypotheses	42
Chapter 2: Materials & Methods	
2.1 Cell culture	
2.1.1 Cell lines	
2.1.2 Cell maintenance	_
2.1.3 Cryopreservation of cells	
2.1.4 Reviving cryopreserved cells	
2.2 RNA/miRNA studies	
2.2.2 Reverse transcription & cDNA synthesis	
2.2.3 Quantitative real-time PCR	
2.2.4 Quantitative real-time PCR analysis	
2.2.5 miRNA microarrays	
2.2.6 3'UTR dual luciferase reporter assay	

	nd miRNA
mimics	
2.2.8 Transient reverse transfection of miRNA mimics	51
2.3 Analysis of Protein Expression	52
2.3.1 Protein extraction & quantitation	52
2.3.2 SDS-PAGE & western blotting	52
2.3.3 Flow cytometry	54
2.4 In vitro functional assays	55
2.4.1 Cell proliferation assay	55
2.4.2 Cell adhesion assay	
2.4.3 Transwell migration assay	
2.4.4 Statistical Analysis	57
Chapter 3: Post-transcriptional regulation of CD151 in prostate	cancer 58
3.1 General introduction	
3.2 Results	
3.2.1 Characterisation of CD151 expression in prostate cells	
3.2.2 Many miRNAs are predicted to regulate the CD151 3'UTR	
3.2.3 Regulation of the CD151 3'UTR in prostate cells	
3.2.4 miRNA expression profiling in prostate cells	
3.2.5 miRNAs that are predicted to regulate CD151 display expression	profiles which
mimic effects of CD151 3'UTR regulation	71
3.2.6 miRNAs bind to the CD151 3'UTR and modulate protein expressi	on72
3.2.7 miR-637 decreases CD151 protein expression in prostate cell line	
3.2.8 miR-637 is predicted to regulate genes involved in cancer associate	•
3.3 Discussion	81
Chapter 4: Post-transcriptional regulation of CD9 in prostate car	ıcer88
Chapter 4: Post-transcriptional regulation of CD9 in prostate car 4.1 General introduction	
	89
4.1 General introduction	91 91 state cancer
4.1 General introduction	91 state cancer 91
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/pr	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg 3'UTR	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to regaling a control of the CD9 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to rega'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but no	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but no	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to rega'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but no	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to rega'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but not contain the contained of the contained contained to the contained contained to the contained contained to the contained contain	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to rega'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but not contain the contained of the contained	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to regalignment of the CD9 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but not consider the constant of the CD9 in brother 5: Post-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in brother 5: Dest-transcriptional Regulation of CD151 & CD9 in Brother 5: Dest-transcriptional Regulation of CD151 & CD9 in Brother 5: Dest-transcriptional Regulation of CD151 & CD9 in Brother 5: Dest-transcriptional Regulation of CD151 & CD9 in Brother 5: Dest-transcriptional Regulation of CD151 & CD9 in Brother 5: Dest-transcriptional Regulation of CD151 & CD9 in Brother 5: Dest-transcriptional Regulation of CD151 & CD9 in Brother 5: Dest-tra	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but not contact the contact of	
4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to reg 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but not considered to the consideration of CD151 & CD9 in brown	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to regalive across a panel of prostate cancer are greated to regalive across a panel of prostate cancer across a panel of prostate cancer across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to regalive across a panel of prostate across a panel of prostate/	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to regature of the CD9 3'UTR 4.2.4 Many miRNAs that are differentially expressed in prostate cancer CD9 3'UTR 4.2.5 miR-518f* decreases CD9 protein levels in prostate cell lines 4.2.6 miR-518f* is predicted to target a range of genes involved in cancer pathways 4.2.7 miR-518f* influences prostate cell migration and adhesion but not contain the contained of the CD151 & CD9 in brown of the CD151 & CD9 in brown of the CD151 & CD9 3'UTR in breast cell lines 5.2.3 Regulation of the CD151 & CD9 3'UTR in breast cell lines 5.2.4 miRNA profiling of human breast cell lines	
4.1 General introduction 4.2 Results 4.2.1 Characterisation of CD9 expression in non-tumourigenic and procells 4.2.2 Regulation of the CD9 3'UTR varies across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to regalive across a panel of prostate cancer are greated to regalive across a panel of prostate cancer across a panel of prostate cancer across a panel of prostate/prcells 4.2.3 Many miRNAs that are deregulated in cancer are predicted to regalive across a panel of prostate across a panel of prostate/	

	reast and
triple negative breast cancer cells	133
5.2.7 miR-518f* modulates migration of non-tumourigenic breast and breast	east cancer
cells	136
5.3 Discussion	141
5.3 Discussion	149
Chapter 7: References	160
5.2.7 miR-518f* modulates migration of non-tumourigenic breast and breast cancells	
•	176

List of figures

Figure 1.1 Schematics of breast and prostate adenocarcinoma progression
Figure 1.2. The general structure of Tetraspanins
Figure 1.3. Tetraspanin Protein Interactions in the Plasma Membrane
Figure 1.4. Tetraspanin-Enriched Microdomains
Figure 1.5. miRNA Biogenesis Pathway
Figure 1.6. Tumour Suppressive and Oncogenic actions of miRNA
Figure 3.1. CD151 profiling of non-tumourigenic prostate and prostate cancer cell lines
Figure 3.2. Inverse correlation between CD151 mRNA and total protein levels in a panel of prostate cell lines
Figure 3.3. CD151 3'UTR mediated regulation in a panel of prostate cell lines
Figure 3.5. qPCR validation of miRNAs differentially expressed in prostate cancer
Figure 3.6. miR-7-2*, miR-637 and miR-619 have increased expression in prostate cells showing CD151 3'UTR repression
Figure 3.7. miR-128, miR-1285 and miR-5480 are highly expressed in prostate cells displaying a high level of CD151 3'UTR repression
Fig 3.8. Many miRNA bind to the CD151 3'UTR and modulate CD151 3'UTR activity
Figure 3.9. miR-637 slightly decreases CD151 protein expression in RWPE1 cells
Figure 3.10. miR-637 and miR-1285 modulate CD151 protein expression in DU145 prostate cancer cells

Figure 4.1. Characterisation of CD9 mRNA and protein expression in prostate cell lines
Figure 4.2. Linear regression analysis of CD9 expression in prostate cell lines
Figure 4.3. Prostate cancer cell lines naturally display repressed CD9 3'UTR activity compared to non-tumourigenic prostate cells
Figure 4.4 miR-106a*, miR-548c-5p, miR-4289 and miR-518f* are upregulated in prostate cells showing changes to CD9 3'UTR activity
Figure 4.5 Many miRNAs predicted to regulate CD9 bind to the CD9 3'UTR in vitro
Figure 4.6. miR-518f* decreases CD9 protein expression in RWPE1 prostate cells
Figure 4.7. miR-518f* decreases CD9 protein expression in DU145 prostate cancer cells
Figure 4.8. miR-518f* increased RWPE1 migration and decreased initial adhesion to ECM substrates
Figure 4.9 miR-518f* significantly decreases DU145 prostate cancer cell migration
Figure 5.1 CD151 expression in a panel of normal breast and breast cancer cell lines
Figure 5.2. Linear regression analysis of CD151 expression in breast cell lines
Figure 5.3 Characterisation of CD9 mRNA and protein expression in normal breast and breast cancer cell lines
Figure 5.4. Analysis of correlations between CD9 mRNA and protein levels in breast cell lines
Figure 5.5 CD151 & CD9 3'UTR activity in a panel of normal breast and breast cancer cell lines
Figure 5.6 Differential expression of miRNAs in breast cancer cells compared to non-tumourigenic breast cells
Figure 5.7 PCR validation of miRNA overexpressed in breast cancer cell lines
Figure 5.8. miR-637 and miR-1226 do not affect CD151 protein expression in breast cell lines

Figure 5.9 Transfection of miR-518f* reduces CD9 protein expression in breast cel lines
Figure 5.10 miR-518f* increases 184A1 cell migration, adhesion and proliferation
Figure 5.11 miR-518f* increases MDA-MB-231 breast cancer cell migration 139
Figure 8.3. Low expression of miR-637 predicts poor survival of breast cance patients

List of tables

Table 1.1. miRNA commonly altered in breast and prostate cancers
Table 2.1. Forward and reverse primers for the detection of mRNA and miRNA
Table 2.2 Table of Antibodies and conditions used for experiments
Table 3.1. miRNA predicted to regulate CD151 and their role in prostate cancer and/or other cancer types
Table 3.2. Analyses of CD151 3'UTR profiles with miRNA microarray data from prostate cell lines
Table 3.3. miR-637 is predicted to regulate genes involved in cancer-associated pathways using miRPath
Table 4.1 miRNAs predicted to bind to and regulate the CD9 3'UTR
Table 4.2 miRNAs with expression levels that are upregulated in prostate cell lines with reduced CD9 3'UTR activity
Table 4.3 miRNAs that are upregulated in prostate cells displaying an Increase or decrease in CD9 3'UTR activity
Table 4.4 Summary of pathways that miR-518f* predicted gene targets are involved in using PANTHER
Table 8.1 Full list of miRNA predicted to regulate the CD151 3'UTR
Table 8.2 Predicted RNA binding motifs in the CD151 3'UTR from RBPmap
Table 8.4 miRNAs predicted to regulate the CD9 3'UTR using a range of target prediction databases
Table 8.5 Predicted RNA binding motifs in the CD9 3'UTR using RBPmap
Table 8.6 miR-518f* is predicted to regulate genes involved in cancer associated pathways

Abstract

Tetraspanins CD151 and CD9 play important roles in cancer progression and metastasis. CD151, referred to as a metastasis enhancer, is typically upregulated in solid malignancies including breast and prostate cancers. In contrast, CD9 is commonly considered a metastasis suppressor, with downregulation of CD9 protein levels in advanced stage cancers. Therefore, CD151 and CD9 are potential targets for new therapeutics to combat cancer progression. However, CD151 and CD9 are not typical "druggable" targets, therefore other ways to change tetraspanin expression such as manipulation of tetraspanin regulation are required. Regulation of CD151 and CD9 expression has been minimally investigated. Therefore, the aim of this thesis was to investigate post-transcriptional regulation of CD151 and CD9 by miRNAs in non-tumourigenic and tumourigenic prostate and breast cell lines.

CD151 mRNA and protein levels were found to inversely correlate in prostate cell lines, with highly tumourigenic prostate cancer cells expressing high levels of CD151 protein. Breast cancer cell lines displayed low levels of CD151 mRNA and protein compared to non-tumuorigenic breast cells, however triple negative MDA-MB-231 breast cancer cells showed similar CD151 protein expression to that of non-tumourigenic breast cells. The degree by which the CD151 3'UTR regulates protein expression was determined with a dual luciferase assay, with greater repression of protein expression found in the tumourigenic cell lines. Bioinformatic analysis of miRNA predicted to bind CD151 together with miRNA expression profiling in prostate cells was used to identify miRNA

that had expression levels matching the luciferase output. miR-637, which was upregulated in prostate cancer cell lines, was shown to regulate expression at the CD151 3'UTR, with transfection of miR-637 mimic into RWPE1 and DU145 prostate cells resulting in a 10-20% decrease in CD151 protein expression. However, miR-637 had no effect on CD151 protein expression in non-tumourigenic 184A1 and MDA-MB-231 breast cancer cells, suggesting a role specifically in prostate cancer.

CD9 mRNA and total protein levels were similar across all prostate cell lines, with typically slightly lower levels of CD9 cell surface levels in tumourigenic cells. In addition breast cancer cell lines displayed lower levels of CD9 mRNA, total protein and cell surface protein expression compared to non-tumourigenic breast cells. A 3'UTR luciferase reporter assay showed that the CD9 3'UTR is differentially regulated in prostate and breast cell lines, with highly tumourigenic prostate cancer cells showing more repression of luciferase compared to other cells. In the panel of breast cells, CD9 3'UTR activity was similar across all lines, however 184A1 breast cells showed increased luciferase, which suggests that the CD9 3'UTR is partly responsible for high CD9 protein levels in normal breast cells. Using the same approach to identifying miRNA as for CD151, miR-518f* was found to bind to the CD9 3'UTR in vitro. Overexpression of miR-518f* in non-tumourigenic prostate RWPE1 and prostate cancer DU145 cells as well as non-tumourigenic breast 184A1 and MDA-MB-231 breast cancer cells led to a significant decrease in CD9 protein expression. Furthermore, transfection of miR-518f* increased migration of RWPE1, 184A1 and MDA-MB-231 cells and decreased migration of DU145 prostate cancer cells. Moreover, overexpression of miR-518f* significantly decreased RWPE1 adhesion to fibronectin

and basement membrane extract and increased 184A1 cell proliferation and adhesion to BME, but had no effect on adhesion in other cell lines or proliferation.

In conclusion, tetraspanins such as CD151 and CD9 are at least partially regulated by miRNAs in prostate and breast cell lines. miRNAs such as miR-518f* may be novel and effective biomarkers and/or therapeutic targets for inhibiting cancer progression in the future.

Publications and conference abstracts arising from this thesis

- D Bond, J Brzozowski, K Skelding, S Roselli & J Weidenhofer. Use of tetraspanins CD151 & CD9 as biomarkers for breast cancer, Breast Cancer Management, March 2014 Vol. 3 issue 2.
- Bond D, Cairns M, Ashman LK & Weidenhofer J. Post-transcriptional regulation of tetraspanins CD151 & CD9 in breast & prostate cancers (poster), AACR 2014 San Diego, USA.
- Bond D, Cairns M, Ashman LK & Weidenhofer J. Post-transcriptional regulation of tetraspanins CD151 and CD9 by micro-RNAs in prostate cancers (oral presentation), Hunter Translational Cancer Conference, Newcastle city hall 2013.
- Bond DR, Cairns MJ, Ashman LK & Weidenhofer J. Post-transcriptional regulation of CD151 and CD9 in prostate and breast cancers (oral presentation), ComBio 9 September – 3 October 2013, Perth, Australia.
- Danielle Bond, Murray Cairns, Leonie K Ashman & Judith Weidenhofer,
 Investigating miRNA Regulation of Tetraspanins CD151 & CD9 in Prostate
 and Breast Cancers (poster presentation), ASMR NSW Scientific Meeting, 3rd
 June 2013, Australian Technology Park Redfern NSW, Australia.
- Danielle Bond, Murray Cairns, Leonie K Ashman & Judith Weidenhofer, Posttranscriptional Regulation of CD151 & CD9 in Prostate Cancers (oral presentation), The 5th Annual Hunter Cancer Research Symposium, 5th November 2012, Newcastle NSW, Australia.
- Danielle Bond, Murray Cairns, Leonie K Ashman & Judith Weidenhofer, Posttranscriptional Regulation of CD151 and CD9 in Breast and Prostate Cancer (oral & poster presentation), 5th European Conference on Tetraspanins, 26-28 September 2012, Nijmegen, the Netherlands.
- Danielle Bond, Murray Cairns, Leonie K Ashman & Judith Weidenhofer,
 Investigating micro-RNA Regulation of Tetraspanins CD151 & CD9 in

Prostate and Breast Cancers (poster), 22nd IUBMB and 37th FEBS Congress: From Single Molecules to Systems Biology, 4th-6th September 2012, Sevilla, Spain.

- Danielle Bond, Matthew J Bowman, Murray Cairns, Leonie K Ashman & Judith Weidenhofer, Investigating Regulation of Tetraspanin Expression in Breast and Prostate Cancers (poster), ASMR NSW Scientific Meeting 2012, Australian Technology Park, Redfern, NSW, Australia.
- Danielle Bond, Murray Cairns, Leonie K Ashman & Judith Weidenhofer,
 Investigating Regulation of Tetraspanin Expression in Prostate Cancer (poster),
 HMRI Cancer Research Symposium, 4th November 2011, Newcastle, NSW,
 Australia.
- Danielle Bond, Murray Cairns, Leonie K Ashman & Judith Weidenhofer,
 Investigating Micro-RNA Regulation of Tetraspanins in Prostate Cancer
 (poster), Australian Society for Medical Research (ASMR) XIX NSW Scientific
 Meeting, 6th June 2011, The University of Sydney, Camperdown, NSW, Australia.
- Danielle Bond, Matthew J Bowman, Murray Cairns, Leonie K Ashman & Judith Weidenhofer, Investigating Regulation of Tetraspanin Expression in Breast and Prostate Cancers (poster), 11th Hunter Cell Biology Meeting, 22-25 March 2011, Hunter Valley Vineyards, NSW, Australia.

Abbreviations

Abbreviation	Word
BC	Breast cancer
BME	Basement membrane extract
cDNA	Complimentary DNA
DCIS	Ductal carcinoma in situ
DNA	Deoxyribonucleic acid
ECM	Extracellular matrix
Exp.	Expression
FN	Fibronectin
miRNA	Micro ribonucleic acid
NTC	Non-targeting control
PC	Prostate cancer
RBP	RNA binding protein
Refs	References
RNA	Ribonucleic acid
TEM	Tetraspanin-enriched microdomain